



Designation: E 394 – 9400

## Standard Test Method for Iron in Trace Quantities Using the 1,10-Phenanthroline Method<sup>1</sup>

This standard is issued under the fixed designation E 394; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon ( $\epsilon$ ) indicates an editorial change since the last revision or reapproval.

### 1. Scope

1.1 This test method covers the determination of iron in the range from 1 to 100  $\mu\text{g}$ .

1.2 This test method is intended to be general for the final steps in the determination of iron and does not include procedures for sample preparation.

1.3 This test method is applicable to samples whose solutions have a pH less than 2. It is assumed that the pH is adjusted to within this range in the sample preparation.

1.4 Review the current material safety data sheets (MSDS) for detailed information concerning toxicity, first-aid procedures, handling, and safety precautions.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

### 2. Referenced Documents

2.1 *ASTM Standards:*

D 1193 Specification for Reagent Water<sup>2</sup>

<sup>1</sup> This test method is under the jurisdiction of ASTM Committee E-15 on Industrial and Specialty Chemicals and is the direct responsibility of Subcommittee E15.201 on Elements—General Standards.

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E 60 Practice for Photometric and Spectrophotometric Methods for Chemical Analysis of Metals<sup>3</sup>

E 180 Practice for Determining the Precision of ASTM Methods for Analysis and Testing of Industrial and Specialty Chemicals<sup>4</sup>

E 200 Practice for Preparation, Standardization, and Storage of Standard and Reagent Solutions for Chemical Analysis<sup>4</sup>

### 3. Summary of Test Method

3.1 This test method is based upon a photometric determination of the 1,10-phenanthroline complex with the iron(II) ion. The sample is dissolved in a suitable solvent and the iron is reduced to the divalent state by the addition of hydroxylamine hydrochloride. The color is then developed, by the addition of 1,10-phenanthroline. After a short reaction period, the absorbance of the solution is measured at approximately 510 nm using a suitable photometer. The absorbance of the solution, once the color is developed, is stable for at least several months.

### 4. Significance and Use

4.1 This test method is suitable for determining trace concentrations of iron in a wide variety of products, provided that appropriate sample preparation has rendered the iron and sample matrix soluble in water or other suitable solvent (see 10.1 and Note 6).

4.2 This test method assumes that the amount of color developed is proportional to the amount of iron in the test solution. The calibration curve is linear over the specified range. Possible interferences are described in Section 5.

### 5. Interferences

5.1 Fortune and Mellon<sup>5</sup> have made a comprehensive study of the interferences of various inorganic ions in this determination. Table 1 and Table 2, taken from their report, show the effects of various cations and anions on the determination of 2.0 ppm iron. If the maximum level of 500 ppm does not interfere, it is very likely that the ion will not interfere in any quantity. The data were obtained under slightly different conditions than those specified in the present test method, but the interferences should be similar. For a more detailed description of interferences, the original literature should be consulted.

<sup>2</sup> Annual Book of ASTM Standards, Vol 11.01.

<sup>3</sup> Annual Book of ASTM Standards, Vol 03.05.

<sup>4</sup> Annual Book of ASTM Standards, Vol 15.05.

<sup>5</sup> Fortune, W. B., and Mellon, M. G., *Industrial and Engineering Chemistry, Analytical Edition*, IENAA Vol 10, 1938, pp. 60–64.

**TABLE 1 Effect of Cations on the Determination of 2 ppm Iron**

Ion	Added As	Maximum Added Without Interference, ppm	Applicable pH Range
Aluminum	AlCl <sub>3</sub>	500	2.0–3.0
Ammonium	NH <sub>4</sub> Cl	500	2.0–9.0
Antimony	SbCl <sub>3</sub>	30	3.0–9.0
Arsenic	As <sub>2</sub> O <sub>5</sub>	500	3.0–9.0
Arsenic	As <sub>2</sub> O <sub>3</sub>	500	3.0–9.0
Barium	BaCl <sub>2</sub>	500	3.0–9.0
Beryllium	Be(NO <sub>3</sub> ) <sub>2</sub>	500	3.0–5.5
Bismuth	Bi(NO <sub>3</sub> ) <sub>3</sub>	... <sup>A</sup>	... <sup>A</sup>
Cadmium	Cd(NO <sub>3</sub> ) <sub>2</sub>	50	3.0–9.0
Calcium	Ca(NO <sub>3</sub> ) <sub>2</sub>	500	2.0–9.0
Chromium	Cr <sub>2</sub> (SO <sub>4</sub> ) <sub>3</sub>	20	2.0–9.0
Cobalt	Co(NO <sub>3</sub> ) <sub>2</sub>	10	3.0–5.0
Copper	Cu(NO <sub>3</sub> ) <sub>2</sub>	10	2.5–4.0
Lead	Pb(C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> ) <sub>2</sub>	500	2.0–9.0
Lithium	LiCl	500	2.0–9.0
Magnesium	Mg(NO <sub>3</sub> ) <sub>2</sub>	500	2.0–9.0
Manganese	MnSO <sub>4</sub>	500	2.0–9.0
Mercury	HgCl <sub>2</sub>	1	2.0–9.0
Mercury	Hg <sub>2</sub> (NO <sub>3</sub> ) <sub>2</sub>	10	3.2–9.0
Molybdenum	(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub>	100	5.5–9.0
Nickel	Ni(NO <sub>3</sub> ) <sub>2</sub>	2	2.5–9.0
Potassium	KCl	1000	2.0–9.0
Silver	AgNO <sub>3</sub>	... <sup>A</sup>	... <sup>A</sup>
Sodium	NaCl	1000	2.0–9.0
Strontium	Sr(NO <sub>3</sub> ) <sub>2</sub>	500	2.0–9.0
Thorium	Th(NO <sub>3</sub> ) <sub>4</sub>	250	2.0–9.0
Tin	H <sub>2</sub> SnCl <sub>6</sub>	20	3.0–6.0
Tin	H <sub>2</sub> SnCl <sub>4</sub>	10	2.0–6.0
Tungsten	Na <sub>2</sub> WO <sub>4</sub>	10	2.5–9.0
Uranium	UO <sub>2</sub> (C <sub>2</sub> H <sub>3</sub> O <sub>2</sub> ) <sub>2</sub>	100	2.0–6.0
Zinc	Zn(NO <sub>3</sub> ) <sub>2</sub>	10	2.0–9.0
Zirconium	Zr(NO <sub>3</sub> ) <sub>4</sub>	50	2.0–9.0

<sup>A</sup>Must be completely absent because of precipitation.

**TABLE 2 Effect of Anions on the Determination of 2 ppm Iron**

Ion	Added As	Maximum Added Without Interference, ppm	Applicable pH Range
Acetate	NaC <sub>2</sub> H <sub>3</sub> O <sub>2</sub>	500	2.0–9.0
Tetraborate	Na <sub>2</sub> B <sub>4</sub> O <sub>7</sub>	500	3.0–9.0
Bromide	NaBr	500	2.0–9.0
Carbonate	Na <sub>2</sub> CO <sub>3</sub>	500	3.0–9.0
Chlorate	KClO <sub>3</sub>	500	2.5–9.0
Chloride	NaCl	1000	2.0–9.0
Citrate	H <sub>3</sub> C <sub>6</sub> H <sub>5</sub> O <sub>7</sub>	500	2.0–9.0
Cyanide	KCN	10	2.0–9.0
Dichromate	K <sub>2</sub> Cr <sub>2</sub> O <sub>7</sub>	20	2.5–9.0
Fluoride	NaF	500	4.0–9.0
Iodide	KI	500	2.0–9.0
Nitrate	KNO <sub>3</sub>	500	2.0–9.0
Nitrite	KNO <sub>2</sub>	500	2.5–9.0
Oxalate	(NH <sub>4</sub> ) <sub>2</sub> C <sub>2</sub> O <sub>4</sub>	500	6.0–9.0
Perchlorate	KClO <sub>4</sub>	100	2.0–9.0
Phosphate	(NH <sub>4</sub> ) <sub>2</sub> HPO <sub>4</sub>	20	2.0–9.0
Pyrophosphate	Na <sub>4</sub> P <sub>2</sub> O <sub>7</sub>	50	6.0–9.0
Silicate	Na <sub>2</sub> SiO <sub>3</sub>	100	2.0–4.5
Sulfate	(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	500	2.0–9.0
Sulfite	Na <sub>2</sub> SO <sub>3</sub>	500	2.0–9.0
Tartrate	(NH <sub>4</sub> ) <sub>2</sub> C <sub>4</sub> H <sub>9</sub> O <sub>6</sub>	500	3.0–9.0
Thiocyanate	KCNS	500	2.0–9.0
Thiosulfate	Na <sub>2</sub> S <sub>2</sub> O <sub>3</sub>	500	3.0–9.0

5.2 Aldehydes, ketones, and oxidizing agents interfere by consuming the hydroxylamine hydrochloride added as a reducing agent.

## 6. Apparatus

6.1 *Photometer*, capable of measuring light absorption at 510 nm and holding a 5-cm or 1-cm cell.

NOTE 1—If a filter photometer is used, a narrow band filter having its maximum transmission at 480 to 520 nm should be used. A discussion of photometers and photometric practice is given in Practice E 60.

6.2 *Absorption Cells*, 5-cm or 1-cm light path.

## 7. Reagents and Materials

7.1 *Purity of Reagents*—Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available.<sup>6</sup> Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

7.2 *Purity of Water*— Unless otherwise indicated, references to water shall be understood to mean Type II reagent water as defined in Specification D 1193.

7.3 *Hydroxylamine Hydrochloride Solution* (100 g/L)—Dissolve 10 g of hydroxylamine hydrochloride (HONH<sub>2</sub>· HCl) in approximately 60 mL of water, filter, and dilute to 100 mL.<sup>7</sup>

7.4 *Iron, Standard Solution* (1 mL = 0.01 mg Fe)<sup>8</sup> (Note 2) —Dissolve 0.1000 g of iron wire in 10 mL of hydrochloric acid (HCl, 1 + 1) and 1 mL of bromine water. Boil until the excess bromine is removed. Add 200 mL of HCl, cool, and dilute to 1 L in a volumetric flask. Dilute 100 mL of this solution to 1 L.

NOTE 2—As an alternative, the standard iron solution may be prepared by weighing exactly 0.7022 g of iron (II) ammonium sulfate hexahydrate (FeSO<sub>4</sub>·(NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>·6H<sub>2</sub>O, minimum purity, 99.5 %) in 500 mL of water containing 20 mL of sulfuric acid (H<sub>2</sub>SO<sub>4</sub>, sp gr 1.84) and diluting to 1 L with water. Dilute 100 mL of this solution to 1 L.

7.5 *1,10-Phenanthroline Solution* (3 g/L)—Dissolve 0.9 g of 1,10-phenanthroline monohydrate in 30 mL of methanol and dilute to 300 mL with water.<sup>7</sup>

7.6 *Ammonium Acetate—Acetic Acid Solution*—Dissolve 100 g of ammonium acetate (CH<sub>3</sub>COONH<sub>4</sub>) in about 600 mL of water, filter, add 200 mL of glacial acetic acid to the filtrate, and dilute to 1 L with water.<sup>7</sup>

## 8. Sampling

8.1 Because this is a general test method for the final steps in determining iron, specific procedures for sample preparation are not included (see 1.3, 4.1 and 4.2).

<sup>6</sup> *Reagent Chemicals, American Chemical Society Specifications*, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopoeial Convention, Inc. (USPC), Rockville, MD.

<sup>7</sup> This solution is also described in Practice E 200.

<sup>8</sup> This solution is used for calibration only.

## 9. Calibration

9.1 By means of suitable pipets or a buret, transfer 0 (reagent blank), 2, 4, 6, 8, and 10 mL, respectively, of the standard iron solution to each of six 100-mL, glass-stoppered graduated cylinders. These cylinders contain 0, 20, 40, 60, 80, and 100 µg of iron, respectively. Dilute the contents of each cylinder to 80 mL with water. Develop the color and measure the absorbance of each calibration standard as described in 10.3 and 10.4.

9.2 Plot, on linear graph paper, the micrograms of iron as a function of absorbance.

NOTE 3—If the photometer readings are percent transmittance, they may be plotted on semi-log paper or converted to absorbance as follows:

$$A = \log \left( \frac{100}{T} \right) \quad (1)$$

where:

$A$  = absorbance, and  
 $T$  = percent transmittance.

NOTE 4—If desired, the slope of the calibration line may be calculated as follows:

$$S = L \times W \quad (2)$$

where:

$S$  = slope of calibration line, µgFe-cm/absorbance unit,  
 $L$  = cell path length, cm, and  
 $W$  = micrograms of iron corresponding to an absorbance of 1.000 on the calibration line.

## 10. Procedure

10.1 Weigh to three significant figures a sample (pH less than 2) containing 1 to 100 µg of iron into a 100-mL, glass-stoppered graduated cylinder (Note 5). If the sample is water soluble, dissolve it in water and dilute to 80 mL with water. If the sample is not water soluble, methanol or another suitable solvent may be used (Note 6).

NOTE 5—The sample size should not exceed 80 mL. When using large samples, the miscibility of the samples and the reagents should be checked before the determination is made. In any case, preliminary tests must be made to determine if the sample or any impurities in the sample interfere in any way with the analysis. If a 1-cm cell is used, the sample must contain at least 5 µg of iron.

NOTE 6—Solvents that have been found suitable for use without recalibration include water, methanol, acetic acid, acetonitrile, and di- and triethylene glycol. Acetone is not suitable. No solvents other than those listed have been tested.

10.2 To prepare a reagent blank, add a quantity of water, approximately equal to the sample size in volume, to a second graduated cylinder. Dilute this to 80 mL with the same solvent used to dissolve the sample.

NOTE 7—When running a number of samples, only one reagent blank is needed. The reagent blank should have the same composition after dilution as the sample. For example, if 10 mL of methanol is taken as a sample, 10 mL of spectro pure methanol should be included in the reagent blank. If 25 mL of methanol is taken as a sample, 25 mL of spectro pure methanol should be included in the reagent blank.

10.3 Add to each cylinder 2 mL of the hydroxylamine hydrochloride solution. Stopper and mix well by inverting the cylinder several times. Add to each cylinder 5 mL of the 1,10-phenanthroline solution and adjust the pH of the solution to between 3.0 and 4.0 by the dropwise addition of the ammonium acetate-acetic acid solution (see Note 8). It may be necessary to adjust the pH of the blank by the addition of dilute HCl. Add to each cylinder 5 mL of the ammonium acetate-acetic acid solution and dilute to 100 mL with water. Stopper and mix well by inverting the cylinder several times. Allow the sample solution and reagent blank to sit at room temperature for a minimum of 15 min.

NOTE 8—It is permissible to prepare the solutions in 150-mL beakers to facilitate the adjustment of the pH using a pH meter. After adjustment, quantitatively transfer the solution to a 100-mL volumetric flask or graduated cylinder for final dilution.

10.4 Measure the absorbance of each sample solution at approximately 510 nm (see Note 9) in a 5-cm cell (see Note 10) using a suitable photometer. Use a matched 5-cm cell filled with the reagent blank to set the instrument at zero absorbance or 100 % transmittance.

NOTE 9—If a filter photometer is used, the same filter should be used for the calibration and sample determinations. When using a spectrophotometer, the wavelength of maximum absorption in the vicinity of 510 nm should be used. This may be determined by scanning the absorption band around 510 nm.

**TABLE 3 Iron Precision**

Level	Repeatability			Within-laboratory, Between-Days			Reproducibility		
	Coefficient of Variation, %	Degrees of Freedom	95 % Limit, %	Coefficient of Variation, %	Degrees of Freedom	95 % Limit, %	Coefficient of Variation, %	Degrees of Freedom	95 % Limit, %
Less than 0.5 ppm	7.540	24	21.11	5.7541	12	16.11	19.2700	4	53.96
Greater than 0.5 ppm	2.7977	14	7.83	3.2222	7	9.02	16.9241	6	47.39

NOTE 10—It is permissible to use matched 1-cm cells for the photometer readings as long as a minimum of 5 µg of iron is present in the sample solution.

10.5 Refer to a previously prepared calibration curve to determine the µg of iron found.

## 11. Calculation

11.1 Calculate the iron content of the sample as follows (Note 11):

$$I_{\text{ron, ppm}} = \frac{B}{W} \quad (3)$$

where:

$B$  = micrograms of iron found, and  
 $W$  = grams of sample.

NOTE 11—If the slope of the calibration line has been calculated, the micrograms of iron,  $B$ , may be calculated as follows:

$$B = A \times \left( \frac{S}{L} \right) \quad (4)$$

where:

$A$  = absorbance of the sample,  
 $B$  = micrograms of iron,  
 $S$  = slope of the calibration line, and  
 $L$  = cell path length, cm.

## 12. Report

12.1 Report the iron content to the nearest 0.01 ppm.

## 13. Precision and Bias

13.1 The following criteria should be used for judging the acceptability of results (see Note 12):

13.1.1 *Repeatability (Single Analyst)*—The coefficient of variation for a single determination has been estimated to be the amount shown in Table 3 at the indicated degrees of freedom. The 95 % limit for the difference between two such runs is the amount shown in Table 3.

13.1.2 *Laboratory Precision (Within-Laboratory, Between-Days)*—The coefficient of variation of results (each the average of duplicates), obtained by the same analyst on different days, has been estimated to be the amount shown in Table 3 at the indicated degrees of freedom. The 95 % limit for the difference between two such averages is the amount shown in Table 3.

13.1.3 *Reproducibility (Multilaboratory)*—The coefficient of variation of results (each the average of duplicates), obtained by analysts on different laboratories, has been estimated to be the amount shown in Table 3 at the indicated degrees of freedom. The 95 % limit for the difference between two such averages is in the amount shown in Table 3.

NOTE 12—The above precision estimates are based on an interlaboratory study performed in 1989 on two samples of water containing approximately 0.5 and 2 ppm iron. One analyst in each of eight laboratories performed duplicate determinations and repeated one day later, for a total of 64 determinations. A second interlaboratory study was performed in 1991 on one sample of ethylene glycol containing approximately 0.2 ppm iron.<sup>9</sup> Practice E 180 was used in developing these precision estimates.

13.2 *Bias*—The bias of this test method has not been determined due to the unavailability of suitable reference materials.

## 14. Keywords

14.1 iron; 1,10-phenanthroline; photometric; spectrophotometric

<sup>9</sup> Details of the interlaboratory study are available from ASTM Headquarters. Request RR: E15-1000.

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